# EPIGENETIC MECHANISMS IN MEMORY FORMATION

# Jonathan M. Levenson and J. David Sweatt

Abstract | Discoveries concerning the molecular mechanisms of cell differentiation and development have dictated the definition of a new sub-discipline of genetics known as epigenetics. Epigenetics refers to a set of self-perpetuating, post-translational modifications of DNA and nuclear proteins that produce lasting alterations in chromatin structure as a direct consequence, and lasting alterations in patterns of gene expression as an indirect consequence. The area of epigenetics is a burgeoning subfield of genetics in which there is considerable enthusiasm driving new discoveries. Neurobiologists have only recently begun to investigate the possible roles of epigenetic mechanisms in behaviour, physiology and neuropathology. Strikingly, the relevant data from the few extant neurobiology-related studies have already indicated a theme — epigenetic mechanisms probably have an important role in synaptic plasticity and memory formation.

Epigenetic mechanisms typically involve heritable alterations in chromatin structure, which, in turn, regulate gene expression. Fundamental insights about epigenetic heritability have come from studies of cell division and development. However, there is increasing evidence that the regulation of chromatin structure through histone acetylation and DNA methylation might mediate longlasting behavioural changes in the context of learning and memory. This idea is fascinating because similar mechanisms are used for triggering and storing long-term memories at the cellular level during, for example, cell differentiation. Another intriguing aspect of this hypothesis is that the storage of lifelong behavioural memory might involve lasting changes in the physical, three-dimensional structure of DNA itself.

Epigenetics is unfamiliar to most neurobiologists. Recently, cellular, molecular and behavioural approaches have led to several exciting developments in this area that specifically concern neurobiological systems. In this review, we first introduce the topic of epigenetics and then discuss the idea that the conservation of epigenetic mechanisms for information storage represents a unifying model in biology, with epigenetic mechanisms being used for cellular memory at different levels that range from cellular differentiation to development to behavioural memory.

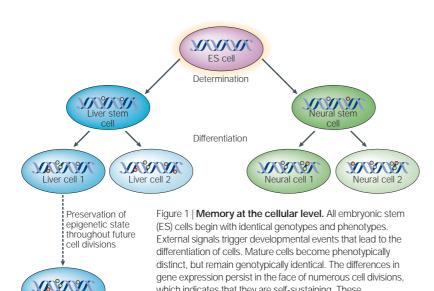
### What is epigenetics?

Epigenetics and its associated terminology have several connotations, and specific terms need to be defined before we can discuss them in detail. We define the genome as a complete set of haploid DNA and the functional units that it encodes. In the nucleus, DNA exists as a highly compressed structure that consists of DNA and protein, known as chromatin. The epigenome is the sum of both the chromatin structure and the pattern of DNA methylation, which is the result of an interaction between the genome and the environment. Three definitions for the term 'epigenetic' are currently in use in the literature.

The broadest definition includes the transmission and perpetuation of information through meiosis or mitosis that is not based on the sequence of DNA. This process is not restricted to DNA-based transmission and can also be protein-based. This definition is broadly used in the yeast literature, wherein phenotypes that can be inherited by daughter cells are perpetuated past cell division using protein-based mechanisms<sup>1-3</sup>.

Developmental biologists and cancer researchers provide a second definition for epigenetic: meiotically and mitotically heritable changes in gene expression that are not coded in the DNA sequence itself. The altered patterns of gene expression can occur through

Department of Neuroscience, Baylor College of Medicine, One Baylor Plaza, S607, Houston, Texas 77030, USA. Correspondence to J.D.S. e-mail: jsweatt@bcm.tmc.edu doi:10.1038/nrn1604 Published online 14 January 2005



several mechanisms that are based on DNA, RNA or proteins (see below)4.

developmentally induced changes in gene expression in mature

cells are mediated by epigenetic regulation of gene expression.

which indicates that they are self-sustaining. These

A third definition posits that epigenetics is the mechanism for the stable maintenance of gene expression that involves physically 'marking' DNA or its associated proteins. This allows genotypically identical cells (such as all cells in an individual human) to be phenotypically distinct (for example, a neuron is phenotypically distinct from a liver cell). The molecular and physical basis for this type of change in DNA or chromatin structure<sup>5</sup> is the focus of this review. By this definition, the regulation of chromatin structure is equivalent to epigenetics.

#### **Epigenetics for information storage**

Several classic examples illustrate the importance of epigenetic mechanisms in information storage at the cellular level. They indicate that epigenetic mechanisms are widely used for the formation and storage of cellular information in response to transient environmental signals. We present these examples to emphasize that the storage of cellular information is in some ways analogous to memory storage in the adult nervous system. Moreover, the lasting cellular changes are triggered by a transient signal in each case, which is also analogous to the formation of behavioural memory in the CNS

A prototype example is mammalian cellular differentiation. Once an embryonic precursor cell is triggered to differentiate into a particular cell type (for example, a liver cell), that cell and its subsequent daughter cells might be required to undergo thousands of cell divisions over the lifetime of the animal. How does a liver cell remember that it is a liver cell when, over the course of cell division, it must replicate de novo its entire genome? The information clearly cannot be contained in the DNA sequence itself. As mentioned above, the answer to this question involves heritable epigenetic mechanisms that allow the cell's identity to be manifest as the subset of genomic DNA that it expresses (FIG. 1).

The genome is marked by, for example, DNA methylation or histone acetylation (or lack thereof) at specific sites that are acquired as part of the differentiation process but are self-perpetuating during DNA replication and cell division. Several hepatic nuclear factors have been shown to be involved in mediating liver-specific gene expression during development through the regulation of histone acetylation<sup>6-8</sup>. Moreover, developmentally-induced changes in histone acetylation are stably propagated from mother to daughter cells in mammals9. So, a liver cell perpetuates its specific acquired pattern of gene expression across cellular generations and over time through these heritable epigenetic marks — an example of lasting memory at the cellular level.

The formation of epigenetic memory is not limited to mammalian cells. Plants are induced to flower by a process known as vernalization that also involves epigenetic mechanisms (for a review, see REF. 10). For example, a biennial plant must experience a period of cold weather between its first and second years of existence for its flowering to be triggered. Exposure to cold in biennial plants results in the activation of epigenetic mechanisms that involve methylation of DNA-binding proteins and acetylation of histones, and these processes trigger mitotically stable changes in the pattern of gene expression. In this way, plant cells 'remember' their exposure to the winter cold and are prepared for the plant to flower during the next spring.

Another example involves T cells of the mammalian immune system. The commitment of T-lymphocyte precursors to a wide variety of differentiated states with different patterns of gene expression is triggered by numerous epigenetic mechanisms that involve DNA methylation and histone modifications (for a review, see REF. 11). These processes are important in the formation of long-lasting immunological memory in response to a transient signal from the environment.

Are these epigenetic mechanisms also extant and operable in non-dividing, terminally differentiated neurons in the adult CNS? Adult neurons no longer have the problem of heritability. However, are the basic epigenetic mechanisms that are important for information storage during development also important for storing memories that manifest themselves behaviourally in the adult? We predict that these mechanisms are conserved in the adult nervous system, where they have been co-opted to serve the formation of behavioural memories. Epigenetic mechanisms subserve changes in neuronal function in the adult that are components of memory at the behavioural level. We propose that epigenetic processes constitute a unified set of molecular mechanisms that allow information storage in systems as diverse as yeast, plants, cellular differentiation and memory storage in the mammalian CNS. To this end, we view chromatin as a dynamic structure that can integrate potentially hundreds of signals from the cell surface and effect a coordinated and appropriate transcriptional response. In the following sections, we explore this hypothesis in more mechanistic detail, focusing on epigenetic mechanisms at the cellular level.

Liver cell

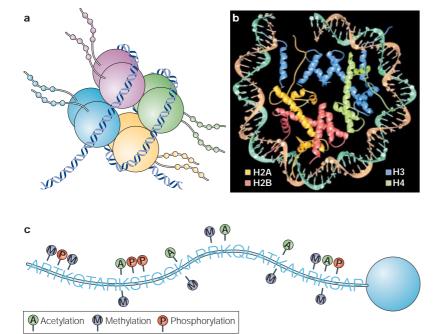


Figure 2 | **The nucleosome and the histone code.** a | Each nucleosome comprises an octamer of histone molecules, which consists of an  ${\rm H3}_2$ – ${\rm H4}_2$  tetramer and two H2A–H2B dimers. The amino (N) termini of histones project out of the nucleosome core and interact with DNA. These histone tails can be epigenetically modified, and function as signal integration platforms. **b** | Crystal structure of the nucleosome depicting the interaction of DNA with histones. Reproduced, with permission, from REF. 12 © (1997) Macmillan Magazines Ltd. **c** | The first 30 amino acids in the N terminus of the human histone H3 are illustrated. Many sites in the N terminus can be targets for epigenetic tagging by, for example, acetylation, phosphorylation and methylation. Regulation of each site is independent, and the integration of epigenetic tags elicits a finely tuned transcriptional response. The integration of signalling at the level of epigenetics is commonly referred to as the histone code 13.

#### Molecular mechanisms

Epigenetic tagging of histones. Histones are highly basic proteins that function to compress DNA within the nucleus and provide a platform for regulating gene transcription. Modification of histones is a mechanism for epigenetic tagging of the genome. Histone modification can occur as a secondary consequence of DNA methylation, or can be mediated by mechanisms that are independent of DNA methylation and controlled by intracellular signalling.

In the nucleus, DNA is tightly packaged into chromatin. The interaction between histone, in the form of the nucleosome (an octamer of histones), and DNA is mediated in part by the amino (N)-terminal tail of histone proteins (FIG. 2a). Structural studies indicate that the N-terminal tails of histones protrude beyond the chromosomes<sup>12</sup> (FIG. 2b). The current hypothesis is that these histone tails serve as signal integration 'platforms', whereby post-translational modifications are combined in a 'histone code' that ultimately directs the activity of numerous transcription factors, cofactors and the transcriptional machinery in general<sup>13</sup> (FIG. 2c). The histone code is the specific pattern of post-translational modifications of a given histone octamer in chromatin. This code, or pattern, is read out as an influence on the specific level of expression of the associated gene(s).

There are several specific sites of post-translational modification within the N-terminal tails of histone proteins, and modifications of these sites modulate the overall structure of chromatin. Currently, four post-translational modifications of histone tails have been characterized: acetylation, methylation, ubiquitylation and phosphorylation, all of which can serve as epigenetic tags.

Acetylation is the best characterized of the post-translational modifications on histones. Acetylation of lysine residues occurs on the amino group in their side chain, which effectively neutralizes their positive charge. The reaction is catalysed by histone acetyltransferases (HATs),which transfer an acetyl group from acetyl-coenzyme A to the  $\epsilon\text{-NH}^+$  group of a Lys residue within a histone  $^{14-17}$ . The process is reversible, and the enzymes that catalyse the reversal of histone acetylation are known as histone deacetylases (HDACs).

Histone methylation — first discovered 40 years ago  $^{18}$  — is another histone-directed epigenetic tag. Similar to acetylation, methylation of histones occurs on  $\epsilon$ -NH $^+$  groups of Lys residues, and is mediated by histone methyltransferases (HMTs). Unlike acetylation, methylation of Lys preserves their positive charge. In addition, Lys can accept up to three methyl groups. Arginine residues within histones can also be mono- or dimethylated on their guanidine nitrogen. This reaction is catalysed by protein Arg methyltransferases (PRMTs).

Ubiquitylation of histones was identified 29 years ago¹¹ but has only recently begun to be characterized in detail. Ubiquitin, a protein of 76 amino acids that is named for its ubiquitous distribution in all cell types and high degree of conservation across species, is usually, but not always, attached to proteins as a signal for degradation by the proteasome²⁰. Similar to other proteins, histones are ubiquitylated through the attachment of a ubiquitin to the  $\epsilon$ -NH⁺ group of a Lys residue²¹. Ubiquitylation of histones H1, H2A, H2B and H3 has been observed¹9,22-2⁴. Most histones seem to be mono-ubiquitylated, although there is evidence for poly-ubiquitylation²¹.

Phosphorylation of histones H1 and H3 was first observed more than 30 years ago in the context of chromosome condensation during mitosis<sup>25,26</sup>. H3 was the first histone whose phosphorylation was characterized in response to the activation of mitogenic signalling pathways<sup>27</sup>. Phosphorylation of serine 10 on H3 is mediated by ribosomal protein S6 kinase 2 (RSK2), which is downstream of extracellular signal-regulated kinase (ERK), mitogen- and stress-activated protein kinase 1 (MSK1), which is downstream of both ERK and mitogen-activated protein kinase 1 (MAPK1 or p38), and the aurora kinase family member increase in ploidy 1 (IPL1) (REFS 28-31). Recent evidence also implicates aurora kinases in the phosphorylation of Ser28 in histone H3 (REF. 32). In order to reverse these phosphorylation events, phosphatases remove phosphate groups from histones<sup>27,33</sup>. So far, the phosphatases PP1 and PP2A have been shown to regulate levels of phorphorylation on H3 (REFS 30,34).

Although a great deal of attention has been given to the N termini of the histones that comprise the nucleosome, increasing evidence indicates that other sites exist for the epigenetic modulation of the genome. Some histones can be modified at domains other than their N termini. For example, disruptor of telomeric silencing 1 protein (DOT1P) has been shown to methylate histone H3 on Lys79, a residue that lies within the globular domain<sup>35</sup>. In addition, higher-order chromatin folding is also undoubtedly involved in the regulation of gene expression. In this regard, there is increasing evidence that the linker histone H1 has a role in the modulation of chromatin structure<sup>36</sup>.

*Epigenetic tagging of DNA.* As mentioned before, the genome can be epigenetically marked by DNA methylation and modifications of histone. Methylation of DNA is catalysed by a class of enzymes known as DNA methyltransferases (DNMTs)37. DNMTs transfer methyl groups to cytosine (C) residues within a continuous stretch of DNA, specifically at the 5-position of the pyrimidine ring<sup>38</sup>. Not all cytosines can be methylated; usually they must be immediately followed by a guanine (G) residue to be methylated<sup>39,40</sup>. These 'CpG' dinucleotide sequences are highly underrepresented in the genome relative to that which would be predicted by chance; however, about 70% of the existing CpG dinucleotides are methylated<sup>41</sup>. The rest of the normally unmethylated CpG dinucleotides occur in small clusters, known as 'CpG islands'42. For purposes of this review, we focus on epigenetic mechanisms that, we speculate, involve reversible DNA methylation, and not the semi-permanent DNA methylation that occurs during cellular differentiation.

DNA methylation leads to marked changes in the structure of chromatin that ultimately result in significant downregulation of transcription. It can directly interfere with the ability of transcription factors to bind to regulatory elements. The transcription factor erythroblastosis 1 (ETS1) and the boundary element CCCTC-binding factor (CTCF) can efficiently bind to non-methylated, but not methylated DNA<sup>43,44</sup>. Moreover, several proteins recognize and bind to methylated CpG residues independent of DNA sequence. The five proteins that are known to bind to methylated CpGs are methyl CpG-binding protein 2 (MECP2), methyl CpG-binding domain protein 1 (MBD1), MBD2, MBD4 and Kaiso<sup>45,46</sup>. These proteins might mediate transcriptional repression by recruiting chromatin-remodelling enzymes. For example, MECP2 directly associates with the transcriptional co-repressor SWI-independent 3A (SIN3A) and histone deacetylase<sup>47,48</sup>.

RNA interference. RNA interference (RNAi) is a mechanism by which the expression of cognate genes is disrupted through the action of double-stranded RNA molecules<sup>49</sup>. Recent studies have indicated that the RNAi machinery is used in the nucleus and is involved in the formation of heterochromatin and epigenetic tagging of histones in yeast. Genetic disruption of the RNAi pathway leads to relaxation of heterochromatin around

centromeres, which causes erroneous expression of normally silent transcripts and a decrease in the methylation of histone H3 (REFS 50,51). Small RNAs that are produced by a specialized ribonuclease might associate with the DNA and direct the formation of a protein complex that promotes the formation of heterochromatin<sup>52</sup>. It is not known whether activity-dependent regulation of neuronal gene expression can be mediated by RNAi. There are a few examples that implicate RNAi in the regulation of gene expression in eukaryotes. The best known example is the expression of the non-coding RNAs XIST (sense) and TSIX (antisense) in X-chromosome inactivation<sup>53,54</sup>. Another example in which RNAi might be involved in regulation of gene expression is in the generation of circadian rhythmicity. In the silkmoth and *Neurospora*, the expression of core clock genes seems to be regulated by endogenous antisense RNA<sup>55,56</sup>. A final example in which RNAi might have a role in the regulation of gene expression involves microRNAs (miRNA). miRNAs are small, non-coding RNAs that have been identified in several metazoans and seem to be involved in development, differentiation and apoptosis<sup>57</sup>.

*Other mechanisms of epigenetic tagging.* We have so far focused on epigenetic mechanisms that are DNAcentric, which result in the modification of either the DNA itself or chromatin structure. According to the broadest definition of epigenetics, which includes any non-DNA-sequence-based system for the perpetuation of information, any protein-based system for the storage of cellular memory is also epigenetic. Prions represent such a viable, protein-based system for epigenetic memory. Once a protein has been converted into its prion form, that protein promotes the transition of other cognate proteins into the prion form. A provocative series of studies has shown that, in Aplysia californica, the cytoplasmic polyadenylation element-binding protein (CPEB) assumes a prion-like conformation after synapses are strengthened<sup>3</sup>. By assuming a prionlike conformation, CPEB can maintain a stable synaptic state in the face of protein turnover (see REFS 2,3 for reviews on protein-based mechanisms of epigenetic tagging).

#### **Epigenetics in neural function**

In the following sections, we explore the functional significance of epigenetics in various aspects of neural function, with an emphasis on chromatin-associated mechanisms. The theme of our discussion will be that synaptic input or other environmental stimuli lead to changes in epigenetic state and ultimately neural function (FIG. 3).

Neural development and differentiation. Neurons express a complement of proteins that are important for their function, but would adversely affect physiological function in other cell types. These include proteins that are involved in excitability, transmitter release and the maintenance of transmembrane potential. Genes that are to be expressed in neurons, but not in other cell

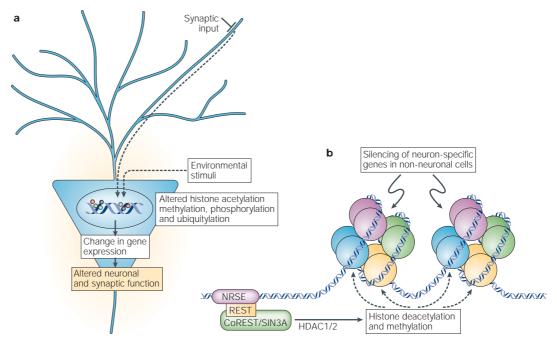


Figure 3 | **Epigenetics in the adult nervous system. a** | Regulation of the epigenetic state of the genome in adult neurons occurs in response to synaptic inputs and/or other environmental stimuli. These external stimuli result in changes in the transcriptional profile of the neuron and, ultimately, neural function. **b** | The RE1-silencing transcription factor (REST)/REST co-repressor (CoREST) system. The neuron-restrictive silencer element (NRSE) is upstream of genes to be silenced in non-neuronal cells and recruits REST as a mediator of transcriptional repression. SIN3A, CoREST and REST function with additional factors such as histone deacetylase 1 (HDAC1) and HDAC2 to lead to chromatin condensation and gene silencing.

types, have a neuron-restrictive silencer element (NRSE) in their promoter  $^{58-60}$ . This regulatory element, which is approximately 21-24 base pairs long, can completely silence a gene in non-neuronal cells  $^{60}$ .

The first step towards understanding how NRSEs confer tissue-specific regulation of gene expression was the identification of the transcription factors that bind to this regulatory element (FIG. 3). The RE1-silencing transcription factor (REST or NRSF) was the first transcription factor shown to bind to NRSEs and repress gene expression<sup>61</sup>. The REST protein is ubiquitously expressed in cells outside the nervous system, where it acts to repress the expression of neuronal genes<sup>61</sup>. Deletion of the REST gene or functional inhibition of the protein in non-neuronal tissues leads to erroneous expression of neuronal genes and embryonic lethality, whereas ectopic expression of REST in the nervous system inhibits expression of neuronal genes, and results in developmental dysfunction<sup>61-63</sup>. Therefore, REST is important in determining whether a cell has a neuronal phenotype.

REST-dependent gene silencing requires the action of transcriptional co-repressors, two of which have been identified as the REST-binding proteins SIN3A and the REST co-repressor (CoREST)<sup>64–66</sup>. The cellular expression pattern of SIN3A is almost identical to that of REST, which indicates that most REST-dependent gene repression might be co-mediated by SIN3A<sup>67</sup>. The expression of CoREST is more restricted, which indicates that it might be important in mediating specific gene expression patterns in subtypes of cell.

REST-mediated gene silencing requires the modulation of chromatin structure. REST/SIN3A repressor complexes are associated with HDAC1, whereas REST/CoREST complexes are associated with HDAC2 (REFS 65,66,68,69). So, REST-dependent gene silencing with either co-repressor seems to involve decreases in histone acetylation. CoREST has also been shown to associate with members of the switch-sucrose non-fermenting (SWI-SNF) complex, which is an ATP-dependent chromatin remodelling complex<sup>70</sup>. Interestingly, REST/ CoREST-dependent chromatin remodelling, including decreases in histone acetylation and increases in DNA methylation, does not seem to be restricted to the immediate region around an NRSE silencer sequence; rather, the formation of heterochromatin extends across several genes that flank an NRSE<sup>71</sup>. These observations indicate that REST-dependent gene silencing, and therefore cellular differentiation, involves the action of several proteins, which, through decreases in histone acetylation and/or increases in DNA methylation, ultimately mark DNA epigenetically for repression.

Circadian rhythmicity. The physiology of most organisms is modulated throughout the day. These daily rhythms persist in the absence of external environmental cues, have a period of approximately 24 hours and are commonly referred to as circadian rhythms. Circadian rhythms are generated endogenously by a biological timekeeping mechanism known as the circadian clock, which comprises intricate feedback loops of transcription and translation<sup>72</sup>. In addition, the

mechanisms that are responsible for entrainment of the circadian clock to the environment rely on signalling mechanisms that induce changes in transcription. In mammals, the master circadian clock resides in the suprachiasmatic nucleus (SCN), which is situated in the anterior hypothalamus<sup>73,74</sup>. Many peripheral tissues have also been shown to have endogenous circadian clocks<sup>74,75</sup>.

The heart of any circadian clock lies in the transcription–translation feedback loop, which could potentially be modulated by epigenetic mechanisms. So, it is possible that the genome undergoes daily changes in its epigenetic state. The acetylation of histones H3 and H4 associated with the promoters of genes that form part of the core molecular clock mechanism are differentially regulated during a circadian cycle<sup>76</sup>. Moreover, infusion of the HDAC inhibitor trichostatin A into the SCN increases the expression of the mouse clock genes period 1 (*Per1*) and *Per2*, which indicates that epigenetic states directly modulate the expression of the molecular components of the circadian clock<sup>76</sup>.

Adjusting the phase of the circadian clock also requires transcription. The most salient phase-resetting environmental stimulus is light, and pulses of light induce changes in the transcription of several genes that comprise the molecular clock<sup>72</sup>. Epigenetic mechanisms seem to be associated with this regulation, as discrete pulses of light induce increases in acetylation of histones H3 and H4 associated with the promoters of *Per1* and Per2 (REF. 76). Moreover, discrete light pulses induce significant increases in the phosphorylation of histone H3 in the SCN in vivo77. These observations indicate that regulation of the epigenetic state of the nucleus is a core molecular mechanism of the circadian clock that is used to generate rhythmic gene expression and to establish a stable phase relationship between gene expression, an animal's behaviour and physiology, and the environment.

Seizures. Different patterns of action potential firing can lead to differential regulation of gene expression. A particularly noteworthy example is seizures, which are widespread bursts of abnormal excitatory synaptic activity in the CNS. Seizures induce many changes in gene expression in the nervous system, which can lead to the development of chronic epilepsy and/or neuro-degeneration. Early studies revealed that transcription of the immediate early gene c-fos was upregulated in many regions of the brain after seizures<sup>78</sup>. Electron microscopy studies showed that c-Fos protein is preferentially localized to the euchromatic regions of chromosomes, which indicates that part of the transcriptional response to seizures involves changes in chromatin structure<sup>79</sup>.

Other studies have shown that expression of the glutamate receptor 2 (GluR2), AMPA ( $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazole propionic acid) receptor subunit and brain-derived neurotrophic factor (BDNF) are also regulated by seizures<sup>80–87</sup>. Expression of GluR2 mRNA is downregulated by seizures, whereas expression of BDNF mRNA is upregulated<sup>82,85–87</sup>. If seizure-induced

changes in gene expression are due to changes in chromatin structure, then changes in chromatin structure should occur around the genes that are known to be regulated. Two important studies have used the chromatin immunoprecipitation (ChIP) approach to monitor post-translational modifications of histones that are located in the promoters of the *BDNF* and *GluR2* genes. In the first study<sup>88</sup>, pilocarpine-induced seizures significantly decreased acetylation of histone H4 in the GluR2 promoter, whereas acetylation of H4 in the P2 promoter of BDNF was significantly increased. In a separate study<sup>89</sup> that modelled a form of human antidepressant therapy, electroconvulsive seizures (ECS) also increased the acetylation of H4 at the P2 promoter of BDNF, and acetylation and phospho-acetylation of H3 were regulated within the P2 and P3 promoters of BDNF. These results indicate that ECS induces complex regulation of the epigenetic state of the BDNF promoters. ECS also had significant effects on the acetylation of H4 and phospho-acetylation of H3 in the c-Fos promoter, and on the acetylation of H3 and H4 in the promoter of cyclic-AMP response-element-binding protein (CREB)89. These data indicate that the synaptic activity or action potential firing that occurs during seizures results in the complex regulation of the epigenetic state of chromatin.

*Memory formation.* In psychological terms, memory describes the processes that are used by the brain for the long-term storage of information. Early studies implicated both transcription and translation as important for the formation of long-term memories<sup>90–94</sup>. Subsequent studies have shown that the formation of long-term memories is a complex process that involves many signalling pathways and the regulation of numerous genes<sup>95–97</sup>.

A recent study has shown that the same processes that lead to the formation of long-term behavioural memories also lead to epigenetic marking of the genome<sup>98</sup>. Contextual fear conditioning is a hippocampusdependent learning model by which an animal learns to associate a novel context with an aversive stimulus<sup>99,100</sup>. Acetylation of histone H3, but not H4, is significantly increased after an animal undergoes contextual fear conditioning98. The formation of long-term contextual fear memories requires NMDA (N-methyl-D-aspartate)-receptor-dependent synaptic transmission and the MEK-ERK/MAPK signalling cascade (where MEK referes to MAPK/ERK kinase) in the hippocampus<sup>101–103</sup>, and inhibition of either of these processes blocks the increase in acetylation of H3 (REF. 98). These observations were the first to indicate that epigenetic tagging of the genome occurs during consolidation of long-term memories. Interestingly, a different form of long-term memory — latent inhibition — has been associated with altered acetylation of H4 but not H3 (REF. 98). This finding indicates that there might be a histone code for memory formation, whereby specific types of memory are associated with specific patterns of histone modification (FIG. 4; BOX 1).

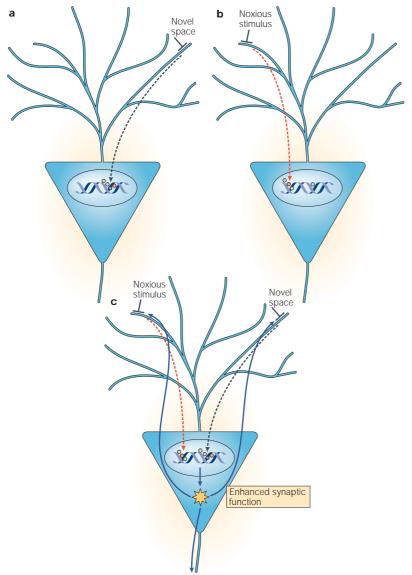


Figure 4 | Model for epigenetics in contextual fear memory — a histone code for memory formation? Exposure of a test subject to various environmental conditions leads to changes in the epigenetic profile of the genome in neurons that reside in relevant brain regions. In this example, we focus on pyramidal neurons in area CA1 of the hippocampus.  $\bf a$  | Exposure of a subject to a novel environment leads to epigenetic changes and formation of novel spatial memories.  $\bf b$  | Exposure to a noxious stimulus leads to epigenetic changes and formation of novel fear memories.  $\bf c$  | Coupling the presentation of the novel environment with the noxious stimulus results in integration of the epigenetic responses, and formation of specific contextual fear memories.

As mentioned above, the addition of acetyl groups to Lys residues in histone proteins is catalysed by HATs. If acetylation of histones were important for the consolidation of long-term memories, then disruption of the activity of HATs would interfere with long-term memory formation. CREB-binding protein (CBP) is a transcriptional co-activator that has endogenous HAT activity  $^{104}$ . Several studies have investigated long-term memory formation in transgenic mice with impaired CBP function. Mice that are heterozygous for a dominant-negative form of truncated CBP (CBP $_{\rm DN}^{\ +/-})^{105}$  have significant deficits in various forms of long-term memory, including step-through passive avoidance,

novel object recognition and cued fear conditioning  $^{105,106}.$  Although these studies provided the first evidence that CBP might be important in long-term memory formation, the wide-ranging developmental abnormalities that are seen in CBP  $_{\rm DN}^{+/-}$  mice make it difficult to interpret the performance of these animals in various memory tasks  $^{105}.$ 

To further determine the role of CBP in long-term memory formation, two recent studies have generated CBP-deficient mice that lack the severe developmental problems of the  $\text{CBP}_{\text{DN}}^{\quad +\!/\!-}$  animals. The first study linked the dominant-negative allele of CBP to an inducible promoter  $(CBP_{I-DN}^{-+/-})^{107}$ . Activation of the dominantnegative allele after animals had developed normally led to impaired learning of the spatial water maze task and novel object recognition<sup>107</sup>. In the second study, mice that lacked one allele of CBP (CBP+/-) had impairments in contextual and cued fear memory, and novel object recognition<sup>108</sup>. In both studies, administration of an HDAC inhibitor restored normal long-term memory formation, which indicates that the balance of HAT/ HDAC activity was altered in these mice and that this caused the memory deficits107,108.

These studies indicate that histone acetylation is regulated by HAT activity and that functional disruption of this process can impair the formation of long-term memories. This might mean that any perturbation in the processes that regulate chromatin structure can affect the formation of long-term memories in vivo. However, can increases in histone acetylation enhance memory formation? To test this directly, two studies investigated the effect of HDAC inhibitors on the formation of long-term memories. Direct infusion of the HDAC inhibitor trichostatin A into the amygdala significantly enhanced the formation of fear-potentiated startle memory<sup>109</sup>. In addition, systemic administration of the HDAC inhibitor sodium butyrate enhanced the formation of contextual fear memories98. In both studies, HDAC inhibitors did not affect short-term memory98,109. In these studies, it was possible that the drugs that were used affected other cellular processes. However, taking all the data into consideration, these studies indicate that long-term behavioural memory regulates, and is regulated by, the epigenome (FIG. 4).

Synaptic plasticity. Synaptic plasticity — activity-dependent changes in synaptic strength — is widely believed to underlie the formation of long-term memories. Many studies have characterized the mechanisms that are responsible for the induction, expression and maintenance of synaptic plasticity in several organisms<sup>110–112</sup>, and one striking observation is that these mechanisms are similar to those that are involved in the formation of long-term memories. So, induction of synaptic plasticity might involve epigenetic mechanisms like those that are involved in long-term memory.

The sensorimotor synapse of the marine mollusc *Aplysia* shows two forms of plasticity. Long-term facilitation (LTF) refers to the lasting enhancement of synaptic transmission, whereas long-term depression (LTD) is a lasting decrease in synaptic transmission.

## Box 1 | Mother's day — every day of your life

Historically, mothers have not been prone to underestimate their lasting impact on their children's behaviour. A recent finding should strengthen their conviction even further 136.

Mouse mothers that show strong nurturing behaviour towards their pups, for example, by frequently licking and grooming their offspring, produce lasting alterations in the patterns of DNA methylation in the CNS of their pups, which apparently persist throughout adulthood <sup>136</sup>. There is evidence that these changes in DNA structure result in decreased anxiety and a strong maternal nurturing instinct in the adult offspring.

Although a detailed review of this landmark study is beyond the scope of this article, this paper is pertinent to the present discussion for several reasons. First, the study indicates that alterations in DNA methylation affect behaviour in the adult. Second, the persistence of neonatally acquired patterns of DNA methylation in the mature CNS is consistent with the hypothesis that epigenetic mechanisms contribute to lasting cellular effects — that is, cellular memory in the CNS. Finally, and perhaps most importantly, the study indicates a specific epigenetic mechanism in the CNS for perpetuating an acquired behavioural characteristic across generations — a particularly robust example of behavioural memory that is potentially subserved by epigenetics.

Acetylation of histone H4 around the promoter of the *Aplysia* CCAAT/enhancer-binding protein (C/EBP) was transiently increased during LTF, but transiently decreased during LTD<sup>113</sup>. Therefore, two opposing forms of plasticity induced opposing changes in histone acetylation in *Aplysia*.

Increases in histone acetylation during LTF in *Aplysia* seem to have functional consequences. Artificial elevation of basal levels of histone acetylation by the HDAC inhibitor trichostatin A transforms short-term facilitation, which does not require the transcription of new genes for its induction, into LTF<sup>113</sup>. So, changes in the overall state of the epigenome can modulate the induction of synaptic plasticity in invertebrates.

Plasticity-induced epigenetic changes are also observed in mammalian models of synaptic plasticity. Long-term potentiation (LTP) is a form of synaptic plasticity whereby synaptic strength is enhanced in response to high-frequency synaptic activity. Several forms of LTP require the activation of NMDA receptors and engagement of the MEK-ERK/MAPK signalling cascade<sup>114-116</sup>. Direct activation of NMDA receptors in the hippocampus leads to an increase in acetylation of histone H3 (REF. 98), which can be blocked by inhibition of the MEK-ERK/MAPK cascade<sup>98</sup>. In addition, activation of dopaminergic, cholinergic and glutamatergic signalling pathways in the hippocampus induces ERKdependent increases in the phosphorylation of histone H3 (REF. 117). These results suggest that the induction of mammalian synaptic plasticity leads to ERK-dependent increases in histone acetylation and phosphorylation in the hippocampus — an interesting parallel to the observations in Aplysia.

Recent studies have directly examined whether histone-modifying enzymes and histone acetylation are necessary for mammalian synaptic plasticity. The induction of early-phase LTP and LTD — forms of plasticity that do not require transcription — was not affected in CBP+/- animals¹08. However, the induction of late-phase LTP, which requires transcription, was significantly impaired in CBP+/- animals¹08. Treatment of hippocampal slices from CBP+/- animals with the HDAC

inhibitor suberoylanilide hydroxamic acid significantly improved late-phase LTP induction, which indicates that inhibition of HDACs had compensated for HAT haploinsufficiency<sup>108</sup>. In other studies using hippocampal slices, induction of LTP using high-frequency stimulation was significantly enhanced by two HDAC inhibitors, trichostatin A and sodium butyrate<sup>98</sup>. In addition, LTP in the amygdala that was induced by forskolin was also enhanced by the HDAC inhibitor trichostatin A<sup>109</sup>. These studies indicate that the epigenetic state of the genome affects the induction of long-term forms of mammalian synaptic plasticity.

# **Epigenetics in human cognition**

There is a considerable body of evidence, albeit indirect, implicating the disruption of epigenetic mechanisms as a causal basis for human cognitive dysfunction. In this section, we briefly describe several instances in which derangements in molecular components of the epigenetic apparatus have been implicated in human cognitive disorders. In interpreting these findings in the present context, an important caveat applies. When considering these cases, it is important to distinguish between a developmental need for epigenetic mechanisms, to allow formation of a normal nervous system, versus an ongoing need for these mechanisms as part of cognitive processing *per se* in the adult. Most of the attention so far has justifiably focused on developmental roles for epigenetics in establishing the capacity for cognitive function in the adult. However, the experimental results outlined above implicate an ongoing and active role for epigenetic mechanisms in memory formation in the adult. So, we believe that it is timely and worthwhile to consider a possible component of cognitive disruption in those disorders outlined below to be due to a loss of the active use of epigenetic mechanisms that are necessary for normal cognition in the mature CNS.

Several disorders of human cognition can be attributed, at least partly, to dysfunction in the mechanisms that underlie epigenetic marking of the genome. Rubinstein–Taybi syndrome (RTS), an inherited autosomal dominant disease, is due to mutations of the transcriptional co-activator, HAT and CBP<sup>118,119</sup>. Several studies using animal models to investigate the molecular basis of RTS indicate that deficiency in CBP has severe consequences for long-term memory formation<sup>105–108</sup>.

Rett syndrome (RS) is an inherited, X-linked disease that seems to be due, at least in part, to a mutation of MECP2 (REFS 120–122). Using animal models, it was discovered that overexpression of MECP2 enhanced long-term memory formation and the induction of hippocampal LTP, indicating that MECP2 modulates memory formation and the induction of synaptic plasticity<sup>123</sup>.

Fragile X syndrome, the most commonly inherited form of mental retardation, is brought about by an abnormal expansion of repeated trinucleotide sequences within one of two Fragile X genes: *FMR1* and *FMR2* (REFS 124,125). *FMR1* and *FMR2* each contain a polymorphic trinucleotide repeat (CGG and CCG,

Table 1 | Epigenetics in human cognitive disorders

lable 1   Epigeneties in Haman cognitive disorders				
Disease	Gene	Function	Epigenetic effect	References
Rubinstein- Taybi Syndrome	CBP	Histone acetyltransferase	↑ histone acetylation	118,119
Rett Syndrome	MECP2	Binds to CpG dinucleotides and recruits HDACs	$\downarrow$ histone acetylation	120–123, 137,138
Fragile X mental retardation	FMR1 and FMR2*	Expansion of CGG or CCG repeats results in aberrant DNA methylation around <i>FMR1</i> and <i>FMR2</i> genes	↑ DNA methylation ↑ histone acetylation	124–127
Alzheimer's disease	APP	APP intracellular domain acts as a Notch-like transcription factor; associated with the HAT TIP60	↓ histone acetylation	128–133
Schizophrenia	reelin	An extracellular matrix protein, involved in synapse development	DNA methylation around the reelin gene	134,135

<sup>\*</sup>Trinucleotide expansions in FMR1 and FMR2. APP, amyloid precursor protein; CBP, cyclic-AMP response-element-binding protein; FMR, fragile X mental retardation; HAT, histone acetyltransferases; HDAC, histone deacetylase; MECP2, methyl CpG-binding protein 2; TIP60, HIV-1 Tat interactive protein, 60kDa.

respectively) in their 5'-untranslated regions that are responsible for the loss of gene expression<sup>126,127</sup>. Expansion of these repeats results in hypermethylation of these regions and flanking CpG islands, leading to transcriptional silencing of the *FMR* and surrounding genes.

The most widespread of senile dementias, Alzheimer's disease, seems to be due, in part, to an increase in soluble  $\beta$ -amyloid peptides in the brain  $^{128}$ . These peptides are created by endo-proteolytic cleavage of the transmembrane amyloid precursor protein (APP) by  $\beta$ - and  $\gamma$ -secretases  $^{129}$ . Interestingly, cleavage of APP results in the production of not only an extracellular  $\beta$ -amyloid fragment, but also an intracellular fragment, the APP intracellular domain (AICD). AICD regulates transcription through recruitment of the adaptor protein FE65 and the HAT TIP60 (HIV-1 Tat interactive protein, 60kDa), which indicates that some of the pathology of Alzheimer's disease is due to misregulation of histone acetylation  $^{130-133}$ .

Finally, schizophrenia is a serious disorder of cognition, rendering sufferers unable to function normally in social situations and in performing everyday cognitive tasks. An emerging body of evidence indicates that deficiencies in the extracellular matrix protein reelin are responsible for the aetiology of schizophrenia<sup>134</sup>. The promoter of reelin contains several sites for DNA

methylation, and inhibitors of HDAC and DNMT activity increase expression of reelin, indicating that epigenetic mechanisms govern the expression of this protein<sup>135</sup>. All these observations indicate that dysfunction of the normal epigenetic status of the genome can have marked consequences on normal cognitive function (TABLE 1). These studies also indicate that drugs that target the epigenome might represent viable therapies for treating various diseases that affect cognition.

#### Conclusions

Chromatin is a dynamic structure that integrates potentially hundreds of signals from the cell surface and effects a coordinated and appropriate transcriptional response. It is increasingly clear that epigenetic marking of chromatin and DNA itself is an important component of the signal integration that is performed by the genome as a whole. Moreover, changes in the epigenetic state of chromatin can have lasting effects on behaviour. We propose that the CNS has co-opted mechanisms of epigenetic tagging of the genome for use in the formation of long-term memories. Moreover, many disorders of human cognition might involve dysfunctions of epigenetic tagging. In our estimation, understanding the epigenetic regulation of neural function will be vital for fully understanding the molecular processes that govern memory formation and human cognition.

- Pray, L. Epigenetics: genome, meet your environment Scientist 18, 14–20 (2004).
  - This is an excellent introduction to the field of epigenetics.
- Sİ, K. et al. A neuronal isoform of CPEB regulates local protein synthesis and stabilizes synapse-specific long-term facilitation in Aplysia. Cell 115, 893–904 (2003).
  - This study shows that the *Aplysia* form of the CPEB protein is necessary for the induction of long-term forms of synaptic plasticity.
- Si, K., Lindquist, S. & Kandel, E. R. A neuronal isoform of the *Aplysia* CPEB has prion-like properties. *Cell* 115, 879–891 (2003).
  - In this study, the authors provide evidence to indicate that the CPEB protein in *Aplysia* contains a prion-like domain that might act in an epigenetic-like manner to mark specific synapses for facilitation.
- Egger, G., Liang, G., Aparicio, A. & Jones, P. A. Epigenetics in human disease and prospects for epigenetic therapy. *Nature* 429, 457–463 (2004).
- Rakyan, V. K., Preis, J., Morgan, H. D. & Whitelaw, E. The marks, mechanisms and memory of epigenetic states in mammals. *Biochem. J.* 356, 1–10 (2001).
  - This is an excellent recent review of epigenetic mechanisms.
- Hatzis, P. & Talianidis, I. Regulatory mechanisms controlling human hepatocyte nuclear factor 4α gene expression. Mol Cell Biol. 21, 7320–7330 (2001).
- Crowe, A. J. et al. Hepatocyte nuclear factor 3 relieves chromatin-mediated repression of the α-fetoprotein gene. J. Biol. Chem. 274, 25113–25120 (1999).
- Parrizas, M. et al. Hepatic nuclear factor 1-α directs nucleosomal hyperacetylation to its tissue-specific transcriptional targets. Mol. Cell. Biol. 21, 3234–3243 (2001).

- Ehrenhofer-Murray, A. E. Chromatin dynamics at DNA replication, transcription and repair. Eur. J. Biochem. 271 2335–2349 (2004)
- Henderson, I. R., Shindo, C. & Dean, C. The need for winter in the switch to flowering. *Annu. Rev. Genet.* 37, 371–392 (2003)
- Smale, S. T. The establishment and maintenance of lymphocyte identity through gene silencing. *Nature Immunol.* 4, 607–615 (2003).
- Luger, K., Mader, A. W., Richmond, R. K., Sargent, D. F. & Richmond, T. J. Crystal structure of the nucleosome core particle at 2.8 Å resolution. *Nature* 389, 251–260 (1997).
- Strahl, B. D. & Allis, C. D. The language of covalent histone modifications. *Nature* 403, 41–45 (2000).
- Tanner, K. G. et al. Catalytic mechanism and function of invariant glutamic acid 173 from the histone acetyltransferase GCN5 transcriptional co-activator. J. Biol. Chem. 274, 18157–18160 (1999).

- Tanner, K. G., Langer, M. R., Kim, Y. & Denu, J. M. Kinetic mechanism of the histone acetyltransferase GCN5 from yeast. J. Biol. Chem. 275, 22048–22055 (2000).
- Lau, O. D. et al. p300/CBP-associated factor histone acetyltransferase processing of a peptide substrate. Kinetic analysis of the catalytic mechanism. J. Biol. Chem. 275, 21953–21959 (2000).
- Tanner, K. G., Langer, M. R. & Denu, J. M. Kinetic mechanism of human histone acetyltransferase P/CAF. *Biochemistry* 39, 11961–11969 (2000).
- Murray, K. The occurrence of ε-N-methyl lysine in histones Biochemistry 127, 10–15 (1964).
- Goldknopf, İ. L. et al. Isolation and characterization of protein A24, a 'histone-like' non-histone chromosomal protein. J. Biol. Chem. 250, 7182–7187 (1975).
- Pickart, C. M. Mechanisms underlying ubiquitination. Annu. Rev. Biochem. 70, 503–533 (2001).
- Nickel, B. E. & Davie, J. R. Structure of polyubiquitinated histone H2A. *Biochemistry* 28, 964–968 (1989).
- histone H2A. *Biochemistry* **28**, 964–968 (1989). 22. West, M. H. & Bonner, W. M. Histone 2B can be modified by the attachment of ubiquitin. *Nucleic Acids Res.* **8**, 4671–4680 (1980).
- Chen, H. Y., Sun, J. M., Zhang, Y., Davie, J. R. & Meistrich, M. L. Ubiquitination of histone H3 in elongating spermatids of rat testes. *J. Biol. Chem.* 273, 13165–13169 (1998).
- Pham, A. D. & Sauer, F. Ubiquitin-activating/conjugating activity of TAFII250, a mediator of activation of gene expression in *Drosophila*. Science 289, 2357–2360 (2000).
- Bradbury, E. M., Inglis, R. J., Matthews, H. R. & Sarner, N. Phosphorylation of very-lysine-rich histone in *Physarum polycephalum*. Correlation with chromosome condensation *Eur. J. Biochem.* 33, 131–139 (1973).
- Gurley, L. R., Walters, R. A. & Tobey, R. A. Cell cycle-specific changes in histone phosphorylation associated with cell proliferation and chromosome condensation. *J. Cell Biol.* 60, 356–364 (1974).
- Mahadevan, L. C., Willis, A. C. & Barratt, M. J. Rapid histone H3 phosphorylation in response to growth factors, phorbol esters, okadaic acid, and protein synthesis inhibitors. *Cell* 65, 775–783 (1991).
- Sassone-Corsi, P. et al. Requirement of Rsk-2 for epidermal growth factor-activated phosphorylation of histone H3. Science 285, 886–891 (1999).
- Thomson, S. et al. The nucleosomal response associated with immediate-early gene induction is mediated via alternative MAP kinase cascades: MSK1 as a potential histone H3/HMG-14 kinase. EMBO J. 18, 4779–4793 (1999)
- Hsu, J. Y. et al. Mitotic phosphorylation of histone H3 is governed by lpl1/aurora kinase and Glc7/PP1 phosphatase in budding yeast and nematodes. Cell 102, 279–291 (2000).
- Di Agostino, S., Rossi, P., Geremia, R. & Sette, C. The MAPK pathway triggers activation of Nek2 during chromosome condensation in mouse spermatocytes. *Development* 129, 1715–1727 (2002).
- Goto, H., Yasui, Y., Nigg, E. A. & Inagaki, M. Aurora-B phosphorylates Histone H3 at serine28 with regard to the mitotic chromosome condensation. *Genes Cells* 7, 11–17 (2002)
- Äjiro, K., Yoda, K., Utsumi, K. & Nishikawa, Y. Alteration of cell cycle-dependent histone phosphorylations by okadaic acid. Induction of mitosis-specific H3 phosphorylation and chromatin condensation in mammalian interphase cells.
   J. Biol. Chem. 271, 13197–13201 (1996).

   Nowak, S. J., Pai, C. Y. & Corces, V. G. Protein phosphatase
- Nowak, S. J., Pai, C. Y. & Corces, V. G. Protein phosphatase 2A activity affects histone H3 phosphorylation and transcription in *Drosophila melanogaster*. Mol. Cell. Biol. 23, 6129–6138 (2003).
- van Leeuwen, F., Gafken, P. R. & Gottschling, D. E. Dot1p modulates silencing in yeast by methylation of the nucleosome core. *Cell* 109, 745–756 (2002).
- Brown, D. T. Histone H1 and the dynamic regulation of chromatin function. *Biochem. Cell Biol.* 81, 221–227 (2003)
- Okano, M., Xie, S. & Li, E. Cloning and characterization of a family of novel mammalian DNA (cytosine-5) methyltransferases. *Nature Genet.* 19, 219–220 (1998).
- Chen, L. et al. Direct identification of the active-site nucleophile in a DNA (cytosine-5)-methyltransferase. Biochemistry 30, 11018–11025 (1991).
- Bird, A. P. Use of restriction enzymes to study eukaryotic DNA methylation: II. The symmetry of methylated sites supports semi-conservative copying of the methylation pattern. J. Mol. Biol. 118, 49–60 (1978).
- Cedar, H., Solage, A., Glaser, G. & Razin, A. Direct detection of methylated cytosine in DNA by use of the restriction enzyme Mspl. *Nucleic Acids Res.* 6, 2125–2132 (1979).
- Cooper, D. N. & Krawczak, M. Cytosine methylation and the fate of CpG dinucleotides in vertebrate genomes. *Hum. Genet.* 83, 181–188 (1989).
- Bird, A. P. CpG-rich islands and the function of DNA methylation. *Nature* 321, 209–213 (1986).

- Maier, H., Colbert, J., Fitzsimmons, D., Clark, D. R. & Hagman, J. Activation of the early B-cell-specific *mb-1* (*ig-α*) gene by Pax-5 is dependent on an unmethylated Ets binding site. *Mol. Cell. Biol.* 23, 1946–1960 (2003).
- Bell, A. C. & Felsenfeld, G. Methylation of a CTCFdependent boundary controls imprinted expression of the *lgf2* gene. *Nature* 405, 482–485 (2000).
- Hendrich, B. & Bird, A. Identification and characterization of a family of mammalian methyl-CpG binding proteins. *Mol. Cell. Biol.* 18, 6538–6547 (1998).
- Prokhortchouk, A. et al. The p120 catenin partner Kaiso is a DNA methylation-dependent transcriptional repressor. Genes Dev. 15, 1613–1618 (2001).
- Jones, P. L. et al. Methylated DNA and MeCP2 recruit histone deacetylase to repress transcription. Nature Genet 19, 187–191 (1998).
- Nan, X. et al. Transcriptional repression by the methyl-CpGbinding protein MeCP2 involves a histone deacetylase complex. Nature 393, 386–389 (1998).
- Montgomery, M. K., Xu, S. & Fire, A. RNA as a target of double-stranded RNA-mediated genetic interference in Caenorhabditis elegans. Proc. Natl Acad. Sci. USA 95, 15502–15507 (1998).
- Volpe, T. A. et al. Regulation of heterochromatic silencing and histone H3 lysine-9 methylation by RNAi. Science 297, 1833–1837 (2002).
- 51. Hall, I. M. et al. Establishment and maintenance of a heterochromatin domain. *Science* **297**, 2232–2237 (2002)
- Verdel, A. et al. RNAi-mediated targeting of heterochromatin by the RITS complex. Science 303, 672–676 (2004).
- by the RITS complex. Science 303, 672–676 (2004).
   Brown, C. J. & Chow, J. C. Beyond sense: the role of antisense RNA in controlling Xist expression. Semin. Cell Dev. Biol. 14, 341–347 (2003).
- Chow, J. C. & Brown, C. J. Forming facultative heterochromatin: silencing of an X chromosome in mammalian females. *Cell. Mol. Life Sci.* 60, 2586–2603 (2003).
- Sauman, I. & Reppert, S. M. Circadian clock neurons in the silkmoth *Antheraea pernyi*: novel mechanisms of Period protein regulation. *Neuron* 17, 889–900 (1996).
- Crosthwalte, S. K. Circadian clocks and natural antisense RNA. FFBS Lett. 567, 49–54 (2004).
- Ambros, V. The functions of animal microRNAs. *Nature* 431, 350–355 (2004).
- Maue, R. A., Kraner, S. D., Goodman, R. H. & Mandel, G. Neuron-specific expression of the rat brain type II sodium channel gene is directed by upstream regulatory elements. *Neuron* 4, 223–231 (1990).
- Li, L., Suzuki, T., Mori, N. & Greengard, P. Identification of a functional silencer element involved in neuron-specific expression of the synapsin I gene. *Proc. Natl Acad. Sci. USA* 90, 1460–1464 (1993).
- Mori, N., Schoenherr, C., Vandenbergh, D. J. & Anderson, D. J. A common silencer element in the SCG10 and type II Na\* channel genes binds a factor present in nonneuronal cells but not in neuronal cells. Neuron 9, 45–54 (1992).
- Chong, J. A. et al. REST: a mammalian silencer protein that restricts sodium channel gene expression to neurons. Cell 80, 949–957 (1995)
- Chen, Z. F., Paquette, A. J. & Anderson, D. J. NRSF/REST is required in vivo for repression of multiple neuronal target genes during embryogenesis. *Nature Genet.* 20, 136–142 (1998).
- Paquette, A. J., Perez, S. E. & Anderson, D. J. Constitutive expression of the neuron-restrictive silencer factor (NRSF)/REST in differentiating neurons disrupts neuronal gene expression and causes axon pathfinding errors in vivo. Proc. Natl Acad. Sci. USA 97, 12318–12323 (2000).
- Andres, M. E. et al. CoREST: a functional co-repressor required for regulation of neural-specific gene expression. Proc. Natl Acad. Sci. USA 96, 9873–9878 (1999).
- Huang, Y., Myers, S. J. & Dingledine, R. Transcriptional repression by REST: recruitment of Sin3A and histone deacetylase to neuronal genes. *Nature Neurosci.* 2, 867–872 (1999).
- Naruse, Y., Aoki, T., Kojima, T. & Mori, N. Neural restrictive silencer factor recruits m5in3 and histone deacetylase complex to repress neuron-specific target genes. *Proc. Natl Acad. Sci. USA* 96, 13691–13696 (1999).
- Grimes, J. A. et al. The co-repressor mSin3A is a functional component of the REST–CoREST repressor complex. J. Biol. Chem. 275, 9461–9467 (2000).
- Roopra, A. et al. Transcriptional repression by neuronrestrictive silencer factor is mediated via the Sin3-histone deacetylase complex. Mol. Cell. Biol. 20, 2147–2157 (2000).
- Ballas, N. et al. Regulation of neuronal traits by a novel transcriptional complex. Neuron 31, 353–365 (2001).
- transcriptional complex. Neuron 31, 353–365 (2001).
  70. Battaglioli, E. et al. REST repression of neuronal genes requires components of the hSWI.SNF complex. J. Biol. Chem. 277, 41038–41045 (2002).

- Lunyak, V. V. et al. Co-repressor-dependent silencing of chromosomal regions encoding neuronal genes. Science 298, 1747–1752 (2002).
- Reppert, S. M. & Weaver, D. R. Coordination of circadian timing in mammals. *Nature* 418, 935–941 (2002).
- Klein, D. C., Moore, R. Y. & Reppert, S. M. Suprachiasmatic Nucleus: The Mind's Clock (Oxford Univ. Press, New York, 1991).
- Zylka, M. J., Shearman, L. P., Weaver, D. R. & Reppert, S. M. Three period homologs in mammals: differential light responses in the suprachiasmatic circadian clock and oscillating transcripts outside of brain. *Neuron* 20, 1103–1110 (1998).
- Balsalobre, A., Damiola, F. & Schibler, U. A serum shock induces circadian gene expression in mammalian tissue culture cells. *Cell* 93, 929–937 (1998).
- Naruse, Y. et al. Circadian and light-induced transcription of clock gene Per1 depends on histone acetylation and deacetylation. Mol. Cell. Biol. 24, 6278–6287 (2004).
- Crosio, C., Cermakian, N., Allis, C. D. & Sassone-Corsi, P. Light induces chromatin modification in cells of the mammalian circadian clock. *Nature Neurosci.* 3, 1241–1247 (2000)
- Morgan, J. I., Cohen, D. R., Hempstead, J. L. & Curran, T. Mapping patterns of c-fos expression in the central nervous system after seizure. *Science* 237, 192–197 (1987).
- Mugnaini, E., Berrebi, A. S., Morgan, J. I. & Curran, T. Fos-like immunoreactivity induced by seizure in mice is specifically associated with euchromatin in neurons. Eur. J. Neurosci. 1, 46–52 (1989).
- Kokaia, M. et al. Suppressed epileptogenesis in BDNF mutant mice. Exp. Neurol. 133, 215–224 (1995).
- Binder, D. K., Routbort, M. J., Ryan, T. E., Yancopoulos, G. D. & McNamara, J. O. Selective inhibition of kindling development by intraventricular administration of TrkB receptor body. J. Neurosci. 19, 1424–1436 (1999).
- Grooms, S. Y., Opitz, T., Bennett, M. V. & Zukin, R. S. Status epilepticus decreases glutamate receptor 2 mRNA and protein expression in hippocampal pyramidal cells before neuronal death. *Proc. Natl Acad. Sci. USA* 97, 3631–3636 (2000).
- Sanchez, R. M. et al. Decreased glutamate receptor 2 expression and enhanced epileptogenesis in immature rat hippocampus after perinatal hypoxia-induced seizures. J. Neurosci. 21, 8154–8163 (2001).
- Isackson, P. J., Huntsman, M. M., Murray, K. D. & Gall, C. M. BDNF mRNA expression is increased in adult rat forebrain after limbic seizures: temporal patterns of induction distinct from NGF. Neuron 6, 937–948 (1991).
- Ernfors, P., Bengzon, J., Kokaia, Z., Persson, H. & Lindvall, O. Increased levels of messenger RNAs for neurotrophic factors in the brain during kindling epileptogenesis. *Neuron* 7, 165–176 (1991).
- Timmusk, T. et al. Multiple promoters direct tissue-specific expression of the rat BDNF gene. Neuron 10, 475–489 (1993).
- Nibuya, M., Morinobu, S. & Duman, R. S. Regulation of BDNF and trkB mRNA in rat brain by chronic electroconvulsive seizure and antidepressant drug treatments. J. Neurosci. 15, 7539–7547 (1995).
- 88. Huang, Y., Doherty, J. J. & Dingledine, R. Altered histone acetylation at glutamate receptor 2 and brain-derived neurotrophic factor genes is an early event triggered by status epilepticus. J. Neurosci. 22, 8422–8428 (2002). Using the pilocarpine model for induction of status epilepticus, the authors show that acetylation of histone H4 is reduced at the promoter for GluR2, but increased at the promoter for the BDNF gene.
- Tsankova, N. M., Kumar, A. & Nestler, E. J. Histone modifications at gene promoter regions in rat hippocampus after acute and chronic electroconvulsive seizures. *J. Neurosci.* 24, 5603–5610 (2004).
  - This paper comprehensively screens the promoter regions of many genes for several types of modification to histones in the context of electroconvulsive shock.
- Barondes, S. H. & Jarvik, M. E. The influence of actinomycin-D on brain RNA synthesis and on memory. J. Neurochem. 11, 187–195 (1964).
- Cohen, H. D. & Barondes, S. H. Further studies of learning and memory after intracerebral actinomycin-D. J. Neurochem. 13, 207–211 (1966).
- Flood, J. F., Bennett, E. L., Orme, E. & Rosenzweig, M. R. Relation of memory formation to controlled amounts of brain protein synthesis. *Physiol. Behav.* 15, 97–102 (1975).
   Flood, J. F., Bennett, E. L., Orme, A. E. & Rosenzweig, M. R.
- Effects of protein synthesis inhibition on memory for active avoidance training. *Physiol. Behav.* **14**, 177–184 (1975). 94. Squire, L. R., Emanuel, C. A., Davis, H. P. & Deutsch, J. A.
- Squire, L. R., Emanuel, C. A., Davis, H. P. & Deutsch, J. A. Inhibitors of cerebral protein synthesis: dissociation of aversive and amnesic effects. *Behav. Biol.* 14, 335–341 (1975).

- Roberson, E. D. & Sweatt, J. D. A biochemical blueprint for long-term memory. *Learn. Mem.* 6, 381–388 (1999).
- Selcher, J. C., Weeber, E. J., Varga, A. W., Sweatt, J. D. & Swank, M. Protein kinase signal transduction cascades in mammalian associative conditioning. *Neuroscientist* 8, 122–131 (2002).
- Levenson, J. M. et al. A bioinformatics analysis of memory consolidation reveals involvement of the transcription factor c-Rel. J. Neurosci. 24, 3933–3943 (2004).
- Levenson, J. M. et al. Regulation of histone acetylation during memory formation in the hippocampus. J. Biol. Chem. 279, 40545–40559 (2004).
  - This study shows that acetylation of H3 is increased by long-term memory formation and activation of the signalling pathways involved in memory formation, and shows that induction of LTP and formation of long-term memories can be enhanced by administration of HDAC inhibitors.
- Phillips, R. G. & LeDoux, J. E. Differential contribution of amygdala and hippocampus to cued and contextual fear conditioning. *Behav. Neurosci.* 106, 274–285 (1992).
- Kim, J. J., Rison, R. A. & Fanselow, M. S. Effects of amygdala, hippocampus, and periaqueductal gray lesions on short- and long-term contextual fear. *Behav. Neurosci.* 107, 1093–1098 (1993).
- Fanselow, M. S., Kim, J. J., Yipp, J. & De Oca, B. Differential effects of the N-methyl-p-aspartate antagonist DL-2-amino-5-phosphonovalerate on acquisition of fear of auditory and contextual cues. *Behav. Neurosci.* 108, 235–240 (1994).
- 102. Atkins, C. M., Selcher, J. C., Petraitis, J. J., Trzaskos, J. M. & Sweatt, J. D. The MAPK cascade is required for mammalian associative learning. *Nature Neurosci.* 1, 602–609 (1998).
- Rampon, C. et al. Enrichment induces structural changes and recovery from nonspatial memory deficits in CA1 NMDAR1-knockout mice. Nature Neurosci. 3, 238–244 (2000)
- Kalkhoven, E. CBP and p300: HATs for different occasions. Biochem. Pharmacol. 68, 1145–1155 (2004).
- 105. Oike, Y. et al. Truncated CBP protein leads to classical Rubinstein–Taybi syndrome phenotypes in mice: implications for a dominant-negative mechanism. Hum. Mol. Genet. 8, 387–396 (1999).
- 106. Bourtchouladze, R. et al. A mouse model of Rubinstein–Taybi syndrome: defective long-term memory is ameliorated by inhibitors of phosphodiesterase 4. Proc. Natl Acad. Sci. USA 100, 10518–10522 (2003).
- Korzus, E., Rosenfeld, M. G. & Mayford, M. CBP histone acetyltransferase activity is a critical component of memory consolidation. *Neuron* 42, 961–972 (2004).
   Using an inducible, dominant-negative form of CBP,
  - Using an inducible, dominant-negative form of CBP, the authors show that derangement of CBP function leads to deficits in long-term memory formation and synaptic plasticity, and that these deficits can be rescued through the use of HDAC inhibitors.
- 108. Alarcon, J. M. et al. Chromatin acetylation, memory, and LTP are impaired in CBP<sup>-/-</sup> mice: a model for the cognitive deficit in Rubinstein–Taybi syndrome and its amelioration. *Neuron* 42, 947–959 (2004).
  - Modelling Rubinstein–Taybi syndrome through CBP haploinsufficiency, this study shows that loss of CBP function leads to deficits in memory formation, and that these deficits can be ameliorated through treatment with an HDAC inhibitor.
- 109. Yeh, S. H., Lin, C. H. & Gean, P. W. Acetylation of nuclear factor-κB in rat amygdala improves long-term but not shortterm retention of fear memory. Mol. Pharmacol. 65, 1286–1292 (2004).

- 110. Malenka, R. C. & Bear, M. F. LTP and LTD; an embarrassment of riches. *Neuron* **44**, 5–21 (2004)
- Klann, E., Antion, M. D., Banko, J. L. & Hou, L. Synaptic plasticity and translation initiation. *Learn. Mem.* 11, 365–372 (2004).
- Pittenger, C. & Kandel, E. R. In search of general mechanisms for long-lasting plasticity: *Aplysia* and the hippocampus. *Philos. Trans. R. Soc. Lond. B* 358, 757–763 (2003).
- Guan, Z. et al. Integration of long-term-memory-related synaptic plasticity involves bidirectional regulation of gene expression and chromatin structure. Cell 111, 483–493 (2002).
  - This study shows that in Aplysia, treatments that induce synaptic facilitation lead to increases in histone acetylation and that treatments that induce synaptic depression lead to decreases in histone acetylation.
- 114. Harris, E. W., Ganong, A. H. & Cotman, C. W. Long-term potentiation in the hippocampus involves activation of N-methyl-p-aspartate receptors. *Brain Res.* 323, 132–137 (1984)
- Morris, R. G., Anderson, E., Lynch, G. S. & Baudry, M. Selective impairment of learning and blockade of long-term potentiation by an N-methyl-p-aspartate receptor antagonist, APS. Nature 319, 774–776 (1986).
- English, J. D. & Sweatt, J. D. A requirement for the mitogenactivated protein kinase cascade in hippocampal long term potentiation. *J. Biol. Chem.* 272, 19103–19106 (1997).
- 117. Crosio, C., Heitz, E., Allis, C. D., Borrelli, E. & Sassone-Corsi, P. Chromatin remodeling and neuronal response: multiple signaling pathways induce specific histone H3 modifications and early gene expression in hippocampal neurons. *J. Cell Sci.* 116, 4905–4914 (2003).
- Petrij, F. et al. Rubinstein-Taybi syndrome caused by mutations in the transcriptional co-activator CBP. Nature 376, 348-351 (1995).
- 119. Blough, R. I. et al. Variation in microdeletions of the cyclic AMP-responsive element-binding protein gene at chromosome band 16p13.3 in the Rublinstein-Taybi syndrome. Am. J. Med. Genet. 90, 29–34 (2000).
- Ellaway, C. & Christodoulou, J. Rett syndrome: clinical characteristics and recent genetic advances. *Disabil. Rehabil.* 23, 98–106 (2001).
- Sirianni, N., Naidu, S., Pereira, J., Pillotto, R. F. & Hoffman, E. P. Rett syndrome: confirmation of X-linked dominant inheritance, and localization of the gene to Xq28. *Am. J. Hum. Genet.* 63, 1552–1558 (1998).
- Amir, R. E. et al. Rett syndrome is caused by mutations in X-linked MECP2, encoding methyl-CpG-binding protein 2. Nature Genet. 23, 185–188 (1999).
- Collins, A. L. et al. Mild overexpression of MeCP2 causes a progressive neurological disorder in mice. Hum. Mol. Genet. 13, 2679–2689 (2004).
- Turner, G., Webb, T., Wake, S. & Robinson, H. Prevalence of fragile X syndrome. Am. J. Med. Genet. 64, 196–197 (1906)
- Ashley, C. T. et al. Human and murine FMR-1: alternative splicing and translational initiation downstream of the CGG-repeat. Nature Genet. 4, 244–251 (1993).
- Gecz, J., Gedeon, A. K., Sutherland, G. R. & Mulley, J. C. Identification of the gene *FMR2*, associated with FRAXE mental retardation. *Nature Genet.* 13, 105–108 (1996).
- 127. Gu, Y., Shen, Y., Gibbs, R. A. & Nelson, D. L. Identification of FMR2, a novel gene associated with the FRAXE CCG repeat and CpG island. Nature Genet. 13, 109–113 (1996).

- Kuo, Y. M. et al. Water-soluble Aβ (N-40, N-42) oligomers in normal and Alzheimer disease brains. J. Biol. Chem. 271, 4077–4081 (1996).
- Selkoe, D. J. The cell biology of β-amyloid precursor protein and presentilin in Alzheimer's disease. *Trends Cell Biol.* 8, 447–453 (1998).
- Sastre, M. et al. Presenilin-dependent γ-secretase processing of β-amyloid precursor protein at a site corresponding to the S3 cleavage of Notch. EMBO Rep. 2, 835–841 (2001).
- 131. Kimberly, W. T., Zheng, J. B., Guenette, S. Y. & Selkoe, D. J. The intracellular domain of the β-amyloid precursor protein is stabilized by Fe65 and translocates to the nucleus in a Notch-like manner. *J. Biol. Chem.* 276, 40288–40292 (2001).
- Cao, X. & Sudhof, T. C. A transcriptionally [correction of transcriptive]] active complex of APP with Fe65 and histone acetyltransferase Tip60. Science 293, 115–120 (2001).
- Von Rotz, R. C. et al. The APP intracellular domain forms nuclear multiprotein complexes and regulates the transcription of its own precursor. J. Cell Sci. 117, 4435–4448 (2004).
- Costa, E. et al. REELIN and schizophrenia: a disease at the interface of the genome and the epigenome. Mol. Intervent. 2, 47–57 (2002).
- 135. Chen, Y., Sharma, R. P., Costa, R. H., Costa, E. & Grayson, D. R. On the epigenetic regulation of the human reelin promoter. *Nucleic Acids Res.* 30, 2930–2939 (2002).
- 136. Weaver, I. C. et al. Epigenetic programming by maternal behavior. Nature Neurosci. 7, 847–854 (2004). This study is one of the first to show that events in early postnatal development result in epigenetic tagging of the genome and can lead to long-term changes in behaviour.
- Shahbazian, M. et al. Mice with truncated MeCP2 recapitulate many Rett syndrome features and display hyperacetylation of histone H3. Neuron 35, 243 (2002)
- Zhao, X. et al. Mice lacking methyl-CpG binding protein 1 have deficits in adult neurogenesis and hippocampal function. Proc. Natl Acad. Sci. USA 100, 6777–6782 (2003)

#### Acknowledgements

The work in the authors' laboratories is supported by the National Institute of Mental Health, National Institute of Child Health and Human Development, National Institute of Neurological Disorders and Stroke, and the American Health Assistance Foundation.

Competing interests statement
The authors declare no competing financial interests.

# Online links

#### DATABASES

# The following terms in this article are linked online to: Entrez Gene:

http://www.ncbi.nih.gov/Entrez/query.fcgi?db=gene APP | BDNF | CTCF | ETS1 | FE65 | FMR1 | FMR2 | GluR2 | HDAC1 | HDAC2 | IPL1 | Kaiso | MAPK1 | MBD1 | MBD2 | MBD4 | MECP2 | MSK1 | Per1 | Per2 | reelin | REST | RSK2 | SIN3A | TIP60 | TSIX | XIST

OMIM: http://www.ncbi.nlm.nih.gov/Omim/ Alzheimer's disease | Rett syndrome | Rubinstein-Taybi syndrome | schizophrenia

Access to this interactive links box is free online.